### ABSTRACT

**C**-Fos is a component of AP-1 transcription factor. Three lines of evidence support pivotal role of c-Fos in learning and memory: (i) learning experience markedly enhances its expression; (ii) blocking od c-Fos impairs, while optogenetic activation of c-Fos expressing neurons supports learning and memory; (iii) c-Fos/AP-1 gene targets in activated neurons, encoding tissue inhibitor of metalloproteinases-1 (TIMP-1) and matrix metalloproteinase 9 (MMP-9) play a major role in synaptic plasticity that underlies learning and memory. TIMP-1 and MMP-9 compose an extracellularly operating enzymatic system active locally around excitatory synapses to modulate their morphology, molecular content and efficacy. Animal studies have implicated MMP-9 in a variety of neuropsychiatric conditions, e.g., epileptogenesis, autism spectrum disorders, development of addiction, and depression. In humans, MMP-9 contributes to epilepsy, alcohol and cocaine addiction, Fragile X Syndrome, schizophrenia and bipolar disorder. In aggregate, all those conditions can be considered as reflecting either healthy or diseased mind.

### C-FOS AS THE FIRST LEARNING-ACTIVATED GENE DISCOVERED

In 1986, after postdoctoral studies on molecular biology of cell proliferation, I joined the Nencki Institute with an idea to initiate investigations into the mind in the brain. My research started with a few assumptions:

- learning and memory offer a window into the mind, and because they can be veridically studied in experimental animals, this is particularly opportune;
- molecular biology provides the most powerful toolbox to approach biological questions;
- cell activation of various kinds follows a partially uniform path, involving enhanced expression of selective genes, such as nuclear proto-oncogenes.

The first two assumptions appear obvious nowadays, so let me comment on the third one. It was discovered in the late 1970s and early 1980s that oncogenes (the genes that give rise to cancer, e.g., via oncogenic retroviruses) have normal, cellular counterpart genes called proto-oncogenes, which are involved in control of the cell division cycle in healthy cells [1,2]. In fact, we demonstrated that the protein c-Myc, encoded by a nuclear proto-oncogene (i.e., protooncogenic protein with the cell nucleus localization), was capable of initiating the cell division cycle [3]. Because c-Myc was found in the cell nuclei, it was immediately hypothesized that this and similarly located protooncogenic proteins (such as c-Fos) might have an impact on gene regulation, thus affecting lasting cellular responses. On other hand, it was shown in the 1960s that inhibitors of de novo protein synthesis, when introduced into the brains of animals, prevented the formation of memories lasting longer than a few hours [4,5]. This led to the conclusion that certain proteins that were produced during the process of learning were crucial for long-term memory. Soon thereafter, it was demonstrated that a similar inhibitory effect on memory was also exerted by substances that block RNA synthesis (i.e. inhibiting gene expression) [5].

Considering those two independent lines of research, a review paper on proto-oncogenes in the cell cycle, proposed that proto-oncogenes might also play a pivotal role in other biological processes thought to involve gene expression [6]. Soon thereafter, the idea suggesting its relevance also to learning and memory was explicitly put forward [7]. Other researchers expressed similar views at that time [8,9]. Therefore, we were encouraged to seek out which genes were activated in the brain under the influence of external stimuli, capable to activate neurons. We initially showed that when glutamic acid, an important neurotransmitter that stimulates neurons to fire and had already been recognized as piv-

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Received: June 15, 2018 Accepted: July 10, 2018

**Key words**: learning and memory; cell adhesion molecules; epilepsy; autism, schizophrenia, addiction

Abbreviations: BA – Basal Amygdala, CaM – cell adhesion molecules, cLTP – chemical LTP, DG – dentate gyrus, ECM – extracellular matrix, FXS – fragile X syndrome, KO – knockout, LTP – long-term potentiation, MMP-9 – matrix metalloproteinase 9, TIMP-1 – tissue inhibitor of metalloproteinases-1 otal for learning and memory, was introduced into the rat brain, it very quickly (within 15 min) and transiently (the effect was gone by 45 min) caused the activation of the gene encoding the c-Fos protein, as measured by the levels of its mRNA [10]. Other studies demonstrated that excessive neuronal activity, such as produced by seizures, also markedly elevated c-Fos expression in the rat brain [11,12], and Hunt *et al.* [13] reported that physiological stimulation of rat primary sensory neurons caused the expression of c-Fos in the spinal cord.

Immediately afterwards, we raised the question of whether evoking long-term potentiation, LTP (that is an electrophysiological model of mechanisms underlying learning and memory) by electrical stimulation in the brain in vivo, as well as exposing a rat to a behavioral, learning episode, might also produce the same effects on gene expression. Indeed that proved to be the case [14-22]. Importantly elevated *c-fos* mRNA expression was observed only when the animals were learning a new task, but not when they vigorously performed already learned behavior [18]. Similar experiments were also carried out by K.V. Anokhin and colleagues, arriving at similar results (activation of the gene encoding c-Fos [23,24]. Over the following years, it has been repeatedly demonstrated that the protein c-Fos appears in nerve cells only when a given neuron is stimulated under conditions making it prone to synaptic plasticity [25] and even specific links of c-Fos expression to long-lasting memories were reported [26].

Despite the wealth of correlative data implicating c-Fos in learning and memory, the functional role of the protein in these phenomena has only recently been proven unequivocally. Using a behavioral paradigm, in which mice had to learn to discriminate sounds of two different frequencies - a task known to rely on functionally intact auditory cortexwe have demonstrated that acquisition of this behavior was greatly impaired when c-Fos was downregulated specifically in this brain area (in result of lentivector-mediated, shRNA-driven blocking of the protein synthesis) [27]. Notably, also auditory discrimination-evoked synaptic plasticity was impaired under those conditions [27]. Synaptic plasticity refers to modifications of synaptic efficacy within vast neuronal network of the brain that processes, in a malleable way, incoming information to produce behaviors that change because of the associated past stimuli and their reinforcing value.

This result is in line with the previous studies on c-Fos knockout and antisense oligonucleotides that negatively affected c-Fos levels [28,29]. However, we shall stress that those previous data suffered from limited specificity, probably because the tools employed diminished basal neuronal activity (that was found intact under our experimental conditions, see de Hoz *et al.* [27]). Most importantly, very elegant studies have conversely shown that artificial, optogenetic stimulation of neurons that expressed c-Fos in response to a learning experience maintain physiological memory traces and even create synthetic ones [30-32]. Thus, it has been found that neurons with transient, learning evoked c-Fos expression are both necessary and sufficient to support the memory trace.

Finally, the last line of evidence pointing to c-Fos role in learning and memory comes from the investigation of its target genes. We have demonstrated that c-Fos activates the gene encoding TIMP-1 (tissue inhibitor of matrix metalloproteinases, Jaworski *et al.* [33]), a protein capable of blocking the activity and thus controlling the extracellular enzyme MMP-9 (matrix metalloptoteinase-9). This, in turn, shifted our attention to MMP-9 as being targeted by TIMP-1. Interestingly, numerous evidence outside the nervous system suggested that also MMP-9 might be regulated by c-Fos/AP-1 [34]. In fact, we have shown that this is the case also in the brain, after behavioral training of fear conditioning and in BDNF-activated neurons in culture [35,36].

### **MMP-9: AN EXTRACELLULAR PROTEASE**

MMP-9, matrix metalloproteinase-9 is a protease that operates predominantly extracellularly. It belongs to a larger family of enzymes (metalloproteinases) that together with other similar molecules form an abundant class of metzincins (including also astacins, ADAMs, ADAMTSs, etc., see for review: [37,38]). Originally, we have reported that MMP-9 undergoes a massive activation in the dentate gyrus (DG) of the hippocampus in response to treatment with kainate [39]. This result was very surprising, since until then multiple studies implicated MMP-9 in pro-neurodegenerative, pathological responses, easily associated with excessive, tissue detrimental proteolysis [40-42]. Although kainate treatment produces CA1 and CA3 hippocampal cell loss, the DG remains spared and even is a subject of massive, albeit aberrant synaptic plasticity [43]. Even more surprising was our discovery that increases in MMP-9 in dendritic tree area of DG concerned not only the protein and enzymatic activity, but also mRNA, suggestive of its translocation towards synapses that were undergoing plastic reorganization. In result, we have put forward hypotheses of possible role of MMP-9 in dendritic remodeling and synaptic plasticity, as well as of local, dendritic/synaptic translation of MMP-9 during plasticity [39].

#### MMP-9 IN SYNAPTIC PLASTICITY, LEARNING AND MEMORY

The hypothesis of an MMP-9 role in physiological plasticity was positively verified by Nagy *et al.* [44] as well as Meighan *et al.* [45] who demonstrated, by various means that MMP-9 was indispensable for late (over half an hour) phase of hippocampal (CA3-CA1) LTP, as well as for hippocampal learning and memory (contextual fear conditioning, water maze), as shown unequivocally with MMP-9 knockout (KO) mice, along other less specific means. Furthermore, increases in MMP-9 protein and enzymatic activity levels under those conditions were also demonstrated [44,45]. Soon thereafter, these observations were extended to other experimental systems of LTP, learning and memory that all involved the hippocampus [46-52]; for review: [53-55].

Surprisingly, formation of aversive memories that relies on lateral amygdala (LaA) apparently does not require MMP-9 activity [44,56], and LTP might be evoked on the external capsule-LaA pathway, even when MMP-9 is missing [57]. An important role of MMP-9 in synaptic function underlying postnatal and even adult cortical plasticity has also been shown for the visual and somatosensory cortex [58-62]. Similarly, requirement for MMP-9 has also been demonstrated for chemical LTP (cLTP) in hippocampal cultures [63-66].

### CENTRAL NUCLEUS OF THE AMYGDALA AS A HUB FOR APPETITIVE PLASTICITY, LEARNING AND MEMORY

Amygdala is a heterogeneous brain structure (at least 13 sub-regions can be easily discerned) that is well known for its pivotal role in emotional and motivational behaviors. Almost 20 years ago we have demonstrated that c-Fos expression in this brain region can be very precisely associated with specific behaviors [67], indicating functional heterogeneity of amygdalar sub-regions. To follow on this finding, we then reported that the central nucleus of the amygdala is strongly c-Fos labeled after appetitive training, but virtually missing c-Fos after acquisition of aversive behaviors in rats and mice [68]. Reviewing all of the existing literature on this topic further reinforced such a notion [69]. Incidentally, we have also found that c-Fos expression in this brain region is conspicuously related to a phenomenon of emotional contagion, a simple form of empathy [70], but is neither associated with acquisition nor extinction of fear behavior [71].

Considering the c-Fos-MMP-9-synaptic plasticity link we have established, we have next investigated whether indeed MMP-9, especially in the central amygdala is critical for appetitive learning and memory. Knapska et al. [56] provided very strong support for such a notion. In particular, we have shown that global KO of MMP-9 impairs appetitive, but not aversive learning and memory and then, selective inhibition of MMP-9 extracellularly within the central amygdala results in the same phenotype - impairs appetitive learning, leaving intact aversive learning and memory [56]. Interestingly, LTP from Lateral Amygdala to Basal Amygdala (BA) and from BA to medial Central Amygdala are greatly impaired when MMP-9 activity is not available [57]. Thus, MMP-9 is not universally mandatory for synaptic plasticity, learning and memory. Nevertheless, activity of this molecule is an obligatory component for specific forms of those phenomena, especially in the hippocampus and central amygdala.

Most recently, Lebitko and colleagues (submitted) have found that selective blocking of c-Fos in the central amygdala by lentivirally delivered specific shRNA (used previously by de Hoz *et al.*, 2018 [27]) impairs specifically preparatory but not consummatory appetitive behaviors. These results go against a mainstream view of central amygdala as playing important role solely in aversive learning, acting as a relay station towards the brain stem. On the other hand, recent data from Tonegawa laboratory have supported our results [72,73], appreciably acknowledging our pioneering findings.

### LOCAL TRANSLATION OF MMP-9

We have also followed the hypothesis of local translation of MMP-9 at/around activated excitatory synapses. It should be noted that our other studies clearly pointed to MMP-9 presence on dendritic spines that harbor postsynaptic areas of excitatory synapses. On other hand, to MMP-9 remained non-detectable at either presynaptic domains or GABA-ergic synapses [74,75]. Konopacki et al. [76], using fluorescent in situ hybridization combined with immunofluorescent protein detection, reported on a patchy MMP-9 mRNA accumulation in DG dendrites in response to kainate treatment. This result reinforced the idea of MMP-9 mRNA being translocated, after kainate, towards excitatory synapses. Dziembowska et al. [77] and Janusz et al. [78] provided a number of experimental data clearly showing that indeed MMP-9 mRNA can undergo local synaptic translation to produce the protein after activation of excitatory synapses. These experiments have also revealed that MMP-9 production, release and synaptic availability after synaptic activation occurs within a few minutes following treatment with glutamate (see also [79]). Importantly, our results have been confirmed and even extended to human brain tissue [80]. Incidentally, another interesting layer of MMP-9 regulation at the mRNA stability level has recently been revealed by Zybura-Broda et al. [81].

### MMP-9 IN STRUCTURAL AND FUNCTIONAL SYNAPTIC PLASTICITY – THE MECHANISMS

The evidence for pivotal role of MMP-9 in structural plasticity of dendritic spines comes from hippocampal cultures and slices, as well as the brain in vivo (see [55,82,83] for review). Two major observations have been made. First, excessive abundance of MMP-9 produces elongation and thinning of the spines [84,85]. On the other hand, physiologically and locally available MMP-9 evokes conversion of small spines to larger, more efficacious mushroom ones [66,86]. We have recently explained this apparent paradox, by finding that the full function of MMP-9 requires first its activity, followed by subsequent inhibition, exerted physiologically by TIMP-1 [87]. We have also found that excessive MMP-9 in transgenic rats with neuronal overexpression of the enzyme [74] results in higher, than in the wild-type rats, proportion of silent synapses and lower AMPA/NMDA receptor ratio, along with impaired LTP. Treatment with MMP inhibitors in those transgenics normalized (i.e., enhanced) LTP as well as unsilenced the synapses, and finally resulted in increased AMPA/NMDA receptor ratio [87].

Using hippocampal cultures subjected to cLTP we have found that this form of synaptic plasticity correlates with growth of small spines into larger mushroom ones, concomitantly with synaptic accumulation of GluA1 AMPA receptors that are at same time less mobile at the synapses [66]. All these major attributes of synaptic plasticity were lost when cLTP treatment was carried under MMP inhibition, i.e., neither spine growth, nor GluA1 accumulation and immobilization at the synapses could be observed under such conditions [66].

Several other reports supported important role of MMP-9 in structural plasticity of dendritic spines, also *in vivo*. Sidhu *et al.* [88] observed that MMP-9 KO mice displayed larger spine head areas in the hippocampus at 1-2 weeks postnatally (at later times this value became the same as in the wild type animals). Interestingly, Aujla and Huntley [89] found that levels of MMP-9 peaked in the hippocampus more or less at the same time. Murase et al. [90] showed that MMP-9 KO mice had unaltered spine density in the hippocampus of adult animals, however, there was increase in proportion of mushroom spine on the expense of thin ones. Kelly *et al.* [62], while studying ocular dominance plasticity in the mouse visual cortex, observed no change in the morphology of existing dendritic spines in MMP-9 KO, however, spine dynamics were altered and KO mice showed increased turnover of dendritic spines over a period of 2 days. Fragkouli et al. [91] constructed mice overexpressing MMP-9 and reported on increased spine density in the hippocampus and somatosensory cortex after behavioral training of adult animals.

# SYNAPTIC CELL ADHESION MOLECULES AS MAJOR NEURONAL MMP-9 TARGETS

It may appear obvious that MMP-9 should cleave components of the extracellular matrix (ECM) surrounding the synapses. In fact, Tsien [92] has proposed that such a cleavage may relieve the synapses/dendritic spines from local environmental constraints limiting their growth, any by this virtue allowing them to undergo plastic changes supporting learning and memory. It should be noted that disruption of ECM may indeed affect synaptic plasticity [93-96]. Although, possible role of MMP-9 in ECM remodeling has been suggested by studies on the cerebellum, no clear MMP-9 substrate has emerged, and in fact we have failed to demonstrate that MMP-9 cleaves suspected substrate, tenascin-C [97,98]. Similarly, it remains as an attractive, though unproven possibility that MMP-9 might cleave CD44 that may anchor hyaluronic acid-based ECM at the neuronal cell membrane [99].

Notably, treatment with excessive exogenous MMP-9 did not produce any gross alteration of ECM in hippocampal cultures [100]. Furthermore, no ECM proteins surrounding synapses have been identified as MMP-9 substrates. In fact, most of such substrates belong to the category of cell adhesion molecules (CAMs, [101-103]). Even more interestingly, all of them are CAMs that are located postsynaptically. The group includes: β-dystroglycan, ICAM-5, neuroligin-1, SynCAM2 (synaptic cell adhesion molecule-2 also known as necl-3) and nectin-3 [79,104-108]. All of those proteins may form trans-synaptic adhesive apparatus with their presynaptic binding partners (β-dystroglycan and neuroligin-1 with neurexins, nectin-3 with nectin-1, ICAM-5 with ICAM-5, SynCAM2 with Syn-Cam1). Other neuronal MMP-9 substrates identified to date are collapsin response mediator protein-2 (CRMP-2, [109]), NGF [110] and pro-BDNF [111].

Considering trans-synaptic adhesive apparatus as a major MMP-9 target and taking into account other aforementioned information, one may suggest that following glutamate stimulation, especially by NMDA receptors, MMP-9 is released from small dendritic spines around postsynaptic domains of excitatory synapses. Next, MMP-9 destabilizes synaptic structure by breaking trans-synaptic connections through limited cleavage of postsynaptically originating proteins bound to their presynaptic partners. This way, the post-synapse and its dendritic spine carrier are allowed to expand and maybe search for a new presynaptic partner. As soon as MMP-9 is inhibited by endogenous TIMP-1, the pre-postsynaptic connection is re-established, however in a modified, possible more efficacious form. Such a scenario is in a perfect agreement with the available experimental data and provides a good explanation for MMP-9 pivotal role in the synaptic plasticity, learning and memory.

However, the molecular mechanisms delineated above, although plausible are not proven yet. Thus, either alternative or complementary modes of MMP-9 function in synaptic plasticity have to be considered. Especially intriguing is partial cleavage of pro-BDNF to produce its mature form [111] as well repeatedly described mediation of MMP-9 synaptic effects via integrins, in particular integrin  $\beta$ 1 [44,63,85,86].

# MMP-9 AS EXECUTOR OF C-FOS FUNCTION IN SYNAPTIC PLASTICITY, LEARNING AND MEMORY?

Considering the aforementioned evidence implicating c-Fos in synaptic plasticity in learning, and its function in regulating MMP-9 and TIMP-1 gene expression, we hypothesize that c-Fos role in these phenomena might in fact be executed via MMP-9 and TIMP-1. A following molecular scenario might be even considered here. During learning experience, glutamate activates NMDA receptors to release MMP-9 and TIMP-1 to control the synaptic plasticity as described above. Since both proteins are released outside the cell and cannot be recuperated, there is a need to replenish them. MMP-9 activity, e.g., by converting pro-BDNF to its mature form (mBDNF) produces a signal that through TrkB receptors and ERK kinases is delivered to SRF transcription factor that is the major upregulator of *c*fos gene expression in activated neurons. Next, the protein product, c-Fos in a form of AP-1, enhances transcription of MMP-9 gene.

### MMP-9 IN NEUROPSYCHIATRIC DISORDERS: A CASE OF ABERRANT SYNAPTIC PLASTICITY?

Besides being pivotal for physiological synaptic plasticity, as described above, MMP-9 has also been implicated in aberrant plasticity that may contribute to a variety of neuropsychiatric conditions [55,112]. A particular strong case has been presented for epileptogenesis (for review: [55,113]), i.e., development of epilepsy. In particular, Wilczynski et al. [74] reported that MMP-9 KO mice were deficient in developing epilepsy in a model of chemical kindling of seizure phenotype, whereas rats overexpressing MMP-9 selectively in neurons were more prone to this phenotype. This genetic proof-of-concept for MMP-9 in epileptogenesis was further supported by studies on chemical inhibitors of the enzyme and other animal models [111,114-116]. Recently, Pijet et al. [117] have employed traumatic brain injury-evoked epileptogenesis, clinically relevant mouse model, to demonstrate important contribution of MMP-9 to epilepsy development. Most interestingly, elevated MMP-9 was found in human epileptic brain samples [118-121]. Furthermore, Zybura-Broda et al. [121] implicated progressive increase in MMP-9 levels, possibly dependent on its gene promoter demethylation, in human epilepsy and rat epileptogenesis.

Similarly, addiction to substances of abuse has repeatedly been linked to MMP-9 levels [122-132]. Our recent study has demonstrated that MMP-9 in the central amygdala controls synaptic plasticity, as well as motivation to seek alcohol in addicted mice and, furthermore, in humans MMP-9 gene polymorphism leading to higher MMP-9 levels supports motivation towards alcohol [132].

Moreover, functional role of MMP-9 has been demonstrated in fragile X syndrome (FXS) that offers a very interesting example of autistic conditions dependent on a single gene (encoding fragile X mental retardation 1 protein, FMRP). Bilousova et al. [84] was the first to show that FMRP KO mice displayed increased MMP-9 activity and then Janusz et al. [78] found that local translation of MMP-9 is negatively controlled by FMRP. Thus, FMRP deficiency releases MMP-9 local translation from this negative control, resulting in higher MMP-9 levels at the synapse. Similarly, Gkogkas et al. [80] found that the eukaryotic translation initiation factor P-eIF4E and MMP-9 expression were both elevated in the brains of human FXS patients and in FMRP deficient mice. Furthermore, Bilousova et al. [84] observed dendritic spine elongation in neuronal cultures that were derived from FMRP KO, a phenomenon that could be normalized by application of minocycline, which inhibited the enzymatic activity of MMP-9. Minocycline treatment also reduced anxiety in FMRP knockout mice [84] and reversed the deficit in ultrasonic vocalizations [133]. Finally, Sidhu et al. [88] crossed MMP-9 KO mice with FMRP KO mice, to alleviate all of the major symptoms of FXS that were observed in FMRP KO. Recently Wen et al. [134] reported that MMP-9 levels were elevated in P12-P18 auditory cortex of Fmr1 KO mice and genetic reduction of MMP-9 to WT levels restored the formation of perineuronal nets (a form of ECM) around parvalbumin expressing inhibitory interneurons. Moreover, in vivo single-unit recordings from auditory cortex neurons showed enhanced spontaneous and sound-driven responses in developing Fmr1 KO mice, which were normalized following genetic reduction of MMP-9. In aggregate, these results addressed abnormal sensory responses associated with FXS and autism spectrum disorders. Notably, the animal data strongly supported the consideration of minocycline as a treatment for FXS. Indeed, several clinical studies that have been conducted, offerring promising results [135-137] (for review, see [139]).

Finally there is also a growing list of findings highly suggestive of a role of MMP-9 in schizophrenia [140] and Lepeta *et al.* [141] have recently shown that MMP-9 gene polymorphism located in 3'UTR of its mRNA contributes to severity of delusional symptoms in humans suffering from schizophrenia, along with affecting local MMP-9 production at the dendritic spines as well as their morphology.

### CONCLUDING REMARKS

The evidence for a role of c-Fos/TIMP-1/MMP-9 pathway in synaptic plasticity appears very compelling, indeed. The evidence derives from studies documenting enhanced c-Fos and MMP-9 levels in response to stimuli that evoke plasticity and, moreover, the MMP-9 increases occur in/ around stimulated excitatory synapses. Furthermore, blocking of c-Fos and MMP-9 impairs the plasticity. These data seem to explain well the role of the c-Fos – MMP-9 pathway in learning and memory, and may be also reveal how MMP-9 contributes to major neuropsychiatric disorders.

#### REFERENCES

- Bishop JM (1990) Nobel Lecture. Retroviruses and oncogenes II. Biosci Rep 10: 473-491
- Varmus HE. Nobel lecture. Retroviruses and oncogenes. I. (1990) Biosci Rep 10: 413-430
- 3. Kaczmarek L, Hyland JK, Watt R, Rosenberg M, Baserga R (1985) Microinjected c-myc as a competence factor. Science 228: 1313-1315
- 4. Davis HP, Squire LR (1984) Protein synthesis and memory: a review. Psychol Bull 96: 518-559
- Matthies H (1989) In search of cellular mechanisms of memory. Prog Neurobiol 32: 77-349
- 6. Kaczmarek L (1986) Protooncogene expression during the cell cycle. Lab Invest 54: 365-376
- Kaczmarek L, Kamińska B (1989) Molecular biology of cell activation. Exp Cell Res 183: 24-35
- Berridge M (1986) Second messenger dualism in neuromodulation and memory. Nature 323: 294-295
- 9. Goelet P, Castellucci VF, Schacher S, Kandel ER (1986) The long and the short of long-term memory-a molecular framework. Nature 322: 419-422
- Kaczmarek L, Siedlecki JA, Danysz W (1988) Protooncogene c-fos induction in rat hippocampus. Mol Brain Res 3: 183-186
- Morgan JI, Cohen DR, Hempstead JL, Curran T (1987) Mapping patterns of c-fos expression in the central nervous system after seizure. Science 237: 192-197
- Dragunow M, Robertson HA (1987) Kindling stimulation induces cfos protein(s) in granule cells of the rat dentate gyrus. Nature 329: 441-442
- Hunt SP, Pini A, Evan G (1987) Induction of c-fos-like protein in spinal cord neurons following sensory stimulation. Nature 328: 632-634
- Kaczmarek L, Nikolajew E (1990) c-fos protooncogene expression and neuronal plasticity. Acta Neurobiol Exp 50: 173-179
- Tishmeyer W, Kaczmarek L, Strausss M, Jork R, Matthies H (1990) Accumulation of c-fos mRNA in rat hippocampus during acquisition of a brightness discrimination. Behav Neural Biol 54: 165-171
- Nikolaev E, Tischmeyer W, Krug M, Matthies H, Kaczmarek L (1991) c-fos protooncogene expression in rat hippocampus and entorhinal cortex following tetanic stimulation of the perforant path. Brain Res 560: 346-349
- Nikolaev E, Kaminska B, Tischmeyer WM, Matthies H, Kaczmarek L (1992) Induction of expression of genes encoding transcription factors in rat brain elicited by behavioral training. Brain Res Bull 28: 479-484
- Nikolaev E, Werka T, Kaczmarek L (1992) c-fos protooncogene expression in rat brain after long term training of two-way active avoidance reaction. Behav Brain Res 48: 91-94.
- Bialy M, Nikolaev E, Beck J, Kaczmarek L (1992) Delayed c-fos expression in sensory cortex following sexual learning in male rats. Mol. Brain Res 14: 352-356
- Kaczmarek L (1992) Expression of c-fos and other genes encoding transcription factors in long term potentiation. Behav Neural Biol 57: 263-266
- Kaczmarek L (1993) Molecular biology of vertebrate learning: is c-fos a new beginning? J Neurosci Res 34: 377-381
- Kaczmarek L (1993) L-glutamate-driven gene expression in learning. Acta Neurobiol Exp 53: 187-196
- Maleeva NE, Ivolgina GL, Anokhin KV, Limborskaia SA (1989) Analysis of the expression of the c-fos proto-oncogene in the rat cerebral cortex during learning. Genetika 25: 1119-1121

- 24. Maleeva NE, Bikbulatova LS, Ivolgina GL, Anokhin KV, Limborskaia SA, Kruglikov RI (1990) Activation of the c-fos proto-oncogene in different structures of the rat brain during training and pseudoconditioning. Dokl Akad Nauk SSSR 31: 762-764
- 25. Kaczmarek L (2002) c-Fos in learning: Beyond the mapping of neuronal activity. In: Handbook of Chemical Neuroanatomy, vol 19: Immediate early genes and inducible transcription factors in mapping of the central nervous system function and dysfunction. Kaczmarek L, Robertson HA (eds) Elsevier, pp 189-216
- 26. Frankland PW, Bontempi B, Talton LE, Kaczmarek L, Silva AJ (2004) The involvement of the anterior cingulate cortex in remote contextual fear memory. Science 304: 881-883
- 27. de Hoz L, Gierej D, Lioudyno V, Jaworski J, Blazejczyk M, Cruces-Solís H, Beroun A, Lebitko T, Nikolaev T, Knapska E, Nelken I, Kaczmarek L. (2018) Blocking c-Fos Expression Reveals the Role of Auditory Cortex Plasticity in Sound Frequency Discrimination Learning. Cereb Cortex 28: 1645-1655
- 28. Fleischmann A, Hvalby O, Jensen V, Strekalova T, Zacher C, Layer LE, Kvello A, Reschke M, Spanagel R, Sprengel R, Wagner EF, Gass P. (2003) Impaired long-term memory and NR2A-type NMDA receptor-dependent synaptic plasticity in mice lacking c-Fos in the CNS. J Neurosci 23: 9116-9122
- 29. Grimm R, Schicknick H, Riede I, Gundelfinger ED, Herdegen T, Zuschratter W, Tischmeyer W (1997) Suppression of c-fos induction in rat brain impairs retention of a brightness discrimination reaction. Learn Mem 3: 402-413
- Garner AR, Rowland DC, Hwang SY, Baumgaertel K, Roth BL, Kentros C, Mayford M (2012) Generation of a synthetic memory trace. Science 335: 1513-1516
- 31. Liu X, Ramirez S, Pang PT, Puryear CB, Govindarajan A, Deisseroth K, Tonegawa S (2012) Optogenetic stimulation of a hippocampal engram activates fear memory recall. Nature 484: 381-385
- 32. Ramirez S, Liu X, Lin PA, Suh J, Pignatelli M, Redondo RL, Ryan TJ, Tonegawa S (2013) Creating a false memory in the hippocampus. Science 341: 387-391
- 33. Jaworski J, Biedermann IW, Lapinska J, Szklarczyk A, Figiel I, Konopka D, Nowicka D, Filipkowski RK, Hetman M, Kowalczyk A, Kaczmarek L (1999) Neuronal excitation-driven and AP-1-dependent activation of timp-1 gene expression in rodent hippocampus. J Biol Chem 274: 28106-28112
- 34. Kaczmarek L, Lapinska-Dzwonek J, Szymczak S (2002) Matrix metalloproteinases in the adult brain physiology: a link between c-Fos, AP-1 and remodeling of neuronal connections? EMBO J 21: 6643-6648
- 35. Ganguly K, Rejmak E, Mikosz M, Nikolaev E, Knapska E, Kaczmarek L (2013) Matrix metalloproteinase (MMP) 9 transcription in mouse brain induced by fear learning. J Biol Chem 288: 20978-20991
- 36. Kuzniewska B, Rejmak E, Malik AR, Jaworski J, Kaczmarek L, Kalita K (2013) Brain-derived neurotrophic factor induces matrix metalloproteinase 9 expression in neurons *via* the serum response factor/c-Fos pathway. Mol Cell Biol 33: 2149-2162
- 37. Rivera S, Khrestchatisky M, Kaczmarek L, Rosenberg GA, Jaworski DM (2010) Metzincin proteases and their inhibitors: foes or friends in nervous system physiology? J Neurosci 30: 15337-15357
- 38. Vandooren J, Van den Steen PE, Opdenakker G (2013) Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9): the next decade. Crit Rev Biochem Mol Biol 48: 222-272
- 39. Szklarczyk A, Lapinska J, Rylski M, McKay RD, Kaczmarek L (2002) Matrix metalloproteinase-9 undergoes expression and activation during dendritic remodeling in adult hippocampus. J Neurosci 22: 920-930
- 40. Yong VW (2005) Metalloproteinases: mediators of pathology and regeneration in the CNS. Nat Rev Neurosci 6: 931-944
- 41. Yong VW, Krekoski CA, Forsyth PA, Bell R, Edwards DR (1998) Matrix metalloproteinases and diseases of the CNS. Trends Neurosci 21: 75-80
- 42. Lo EH, Wang X, Cuzner ML (2002) Extracellular proteolysis in brain injury and inflammation: role for plasminogen activators and matrix metalloproteinases. J Neurosci Res 69: 1-9

- Zagulska-Szymczak S, Filipkowski RK, Kaczmarek L (2001) Kainateinduced genes in the ns from expression patterns. Neurochem Int 38: 485-501
- 44. Nagy V, Bozdagi O, Matynia A, Balcerzyk M, Okulski P, Dzwonek J, Costa RM, Silva AJ, Kaczmarek L, Huntley GW (2006) Matrix metalloproteinase-9 is required for hippocampal late-phase long-term potentiation and memory. J. Neurosci 26: 1923-1934
- 45. Meighan SE, Meighan PC, Choudhury P, Davis CJ, Olson ML, Zornes PA, Wright JW, Harding JW (2006) Effects of extracellular matrix-degrading proteases matrix metalloproteinases 3 and 9 on spatial learning and synaptic plasticity. J Neurochem 96: 1227-1241
- 46. Nagy V, Bozdagi O, Huntley GW (2007) The extracellular protease matrix metalloproteinase-9 is activated by inhibitory avoidance learning and required for long-term memory. Learn Mem 14: 655-664
- Bozdagi O, Nagy V, Kwei KT, Huntley GW (2007) *In vivo* roles for matrix metalloproteinase-9 in mature hippocampal synaptic physiology and plasticity. J Neurophysiol 98: 334-344
- 48. Okulski P, Jay TM, Jaworski J, Duniec K, Dzwonek J, Konopacki FA, Wilczynski GM, Sánchez-Capeli A, Mallet J, Kaczmarek L (2007) TIMP-1 abolishes MMP-9-dependent long-lasting long-term potentiation in the prefrontal cortex. Biol Psychiatry 62: 359-363
- 49. Wright JW, Meighan SE, Murphy ES, Holtfreter KL, Davis CJ, Olson ML, Benoist CC, Muhunthan K. and Harding J. W. (2006) Habituation of the head-shake response induces changes in brain matrix metalloproteinases-3 (MMP-3) and -9. Behav Brain Res 174: 78-85
- 50. Wright JW, Brown TE, Harding JW (2007) Inhibition of hippocampal matrix metalloproteinase-3 and -9 disrupts spatial memory. Neural Plast 2007: 73813
- Conant K, Wang Y, Szklarczyk A, Dudak A, Mattson MP, Lim ST (2010) Matrix metalloproteinase-dependent shedding of intercellular adhesion molecule-5 occurs with long-term potentiation. Neuroscience 166: 508-521
- 52. Wojtowicz T, Mozrzymas JW (2010) Late phase of long-term potentiation in the mossy fiber-CA3 hippocampal pathway is critically dependent on metalloproteinases activity. Hippocampus 20: 917-921
- 53. Huntley GW (2012) Synaptic circuit remodelling by matrix metalloproteinases in health and disease. Nat Rev Neurosci 13: 743-757
- 54. Tsilibary E, Tzinia A, Radenovic L, Stamenkovic V, Lebitko T, Mucha M, Pawlak R, Frischknecht R, Kaczmarek L. (2014) Neural ECM proteases in learning and synaptic plasticity. Prog Brain Res 214: 135-157
- 55. Vafadari B, Salamian A, Kaczmarek L (2016) MMP-9 in translation: from molecule to brain physiology, pathology and therapy. J Neurochem 139 Suppl 2: 91-114
- 56. Knapska E, Lioudyno V, Kiryk A, Mikosz M, Górkiewicz T, Michaluk P, Gawlak M, Chaturvedi M, Mochol G, Balcerzyk M, Wojcik DK, Wilczynski GM, Kaczmarek L (2013) Reward Learning Requires Activity of Matrix Metalloproteinase-9 in the Central Amygdala. J Neurosci 33: 14591-14600
- 57. Gorkiewicz T, Balcerzyk M, Kaczmarek L, Knapska E (2015) Matrix metalloproteinase 9 (MMP-9) is indispensable for long term potentiation in the central and basal but not in the lateral nucleus of the amygdala. Front Cell Neurosci 9: 73
- 58. Szklarczyk A, Kaczmarek L (2005) Physiology of matrix MMPs and their tissue inhibitors in the brain. Biotech Int 17: 15-18
- 59. Spolidoro M, Putignano E, Munafo C, Maffei L, Pizzorusso T (2012) Inhibition of matrix metalloproteinases prevents the potentiation of nondeprived-eye responses after monocular deprivation in juvenile rats. Cereb Cortex 22: 725-734
- 60. Kaliszewska A, Bijata M, Kaczmarek L, Kossut M (2012) Experience-dependent plasticity of the barrel cortex in mice observed with 2-DG brain mapping and c-Fos: effects of MMP-9 KO. Cereb Cortex 22: 2160-2170
- 61. Verslegers M, Lemmens K, Van Hove I, Moons L (2013) Matrix metalloproteinase-2 and -9 as promising benefactors in development, plasticity and repair of the nervous system. Prog Neurobiol 105: 60-78
- 62. Kelly EA, Russo AS, Jackson CD, Lamantia CE, Majewska AK (2015) Proteolytic regulation of synaptic plasticity in the mouse primary visu-

al cortex: analysis of matrix metalloproteinase 9 deficient mice. Front Cell Neurosci 9: 369

- 63. Niedringhaus M, Chen X, Dzakpasu R, Conant K (2012) MMPs and soluble ICAM-5 increase neuronal excitability within *in vitro* networks of hippocampal neurons. PLoS One 7: e42631
- 64. Niedringhaus M, Chen X, Conant K, Dzakpasu R (2013) Synaptic Potentiation Facilitates Memory-like Attractor Dynamics in Cultured *In vitro* Hippocampal Networks. PLoS One 8: e57144
- 65. Szepesi Z, Bijata M, Ruszczycki B, Kaczmarek L, Włodarczyk . (2013) Matrix metalloproteinases regulate the formation of dendritic spine head protrusions during chemically induced long-term potentiation. PLoS One 8: e63314
- 66. Szepesi Z, Hosy E, Ruszczycki B, Bijata M, Pyskaty M, Bikbaev A, Heine M, Choquet D, Kaczmarek L, Włodarczyk J (2014) Synaptically released matrix metalloproteinase activity in control of structural plasticity and the cell surface distribution of GluA1-AMPA receptors. PLoS One 9: e98274
- 67. Savonenko A, Filipkowski RK, Werka T, Zielinski K, Kaczmarek L (1999) Defensive conditioning-related functional heterogeneity among nuclei of rat amygdala revealed by c-Fos mapping. Neuroscience 94: 723-733
- 68. Knapska A, Walasek G, Nikolaev E, Neuhäusser-Wespy F, LippH P, Kaczmarek L, Werka T (2006) Differential involvement of the central amygdala in appetitive versus aversive learning. Learn Mem 13: 192-200
- 69. Knapska E, Radwanska K, Werka T, Kaczmarek L (2007) Functional internal complexity of amygdala: focus on gene activity mapping following behavioral training and drugs of abuse. Physiol Rev 87: 1113-1173
- 70. Knapska E, Nikolaev E, Boguszewski P, Walasek G, Blaszczyk J, Kaczmarek L, Werka T (2006) Between-subject transfer of emotional information evokes specific pattern of amygdala activation. Proc Natl Acad Sci USA 103: 3858-3862
- 71. Knapska E, Macias M, Mikosz M, Nowak A, Owczarek D, Wawrzyniak M, Pieprzyk M, Cymerman IA, Werka T, Sheng M, Maren S, Jaworski J, Kaczmarek L (2012) Functional anatomy of neural circuits regulating fear and extinction. Proc Natl Acad Sci USA 109: 17093-17098
- 72. Kim J, Pignatelli M, Xu S, Itohara S, Tonegawa S (2016) Antagonistic negative and positive neurons of the basolateral amygdala. Nat Neurosci 19: 1636-1646
- 73. Kim J, Zhang X, Muralidhar S, LeBlanc SA, Tonegawa S. (2017) Basolateral to central amygdala neural circuits for appetitive behaviors. Neuron 93: 1464-1479
- 74. Wilczynski GM, Konopacki FA, Wilczek E, Lasiecka Z, Gorlewicz A, Michaluk P, Wawrzyniak M, Malinowska M, Okulski P, Kolodziej LR, Konopka W, Duniec K, Mioduszewska B, Nikolaev E, Walczak A, Owczarek D, Gorecki DC, Zuschratter W, Ottersen OP, Kaczmarek L (2008) Important role of matrix metalloproteinase 9 in epileptogenesis. J Cell Biol 180: 1021-1035
- 75. Gawlak M, Gorkiewicz T, Gorlewicz A, Konopacki FA, Kaczmarek L, Wilczynski GM (2009) High resolution In situ zymography reveals matrix metalloproteinase activity at glutamatergic synapses. Neuroscience 158: 167-176
- 76. Konopacki FA, Rylski M, Wilczek E, Amborska R, Detka D, Kaczmarek L, Wilczyński GM (2007) Synaptic localization of seizure-induced matrix metalloproteinase-9 mRNA. Neuroscience 150: 31-39
- 77. Dziembowska M, Milek J, Janusz A, Rejmak E, Romanowska E, Gorkiewicz T, Tiron A, Bramham CR, Kaczmarek L (2012) Activity-dependent local translation of matrix metalloproteinase-9. J Neurosci 32: 14538-14547
- 78. Janusz A, Milek J, Perycz M, Pacini L, Bagni C, Kaczmarek L, Dziembowska M (2013) The Fragile X mental retardation protein regulates matrix metalloproteinase 9 mRNA at synapses. J Neurosci 33: 18234-18241
- 79. Michaluk P, Kolodziej L, Mioduszewska B, Wilczynski GM, Dzwonek J, Jaworski J, Gorecki DC, Ottersen OP, Kaczmarek L (2007) beta-dys-

troglycan as a target for MMP-9, in response to enhanced neuronal activity. J Biol Chem 282: 16036-16041

- 80. Gkogkas CG, Khoutorsky A, Cao R, Jafarnejad SM, Prager-Khoutorsky M, Giannakas N, Kaminari A, Fragkouli A, Nader K, Price TJ, Konicek BW, Graff JR, Tzinia AK, Lacaille JC, Sonenberg N (2014) Pharmacogenetic inhibition of eIF4E-dependent Mmp9 mRNA translation reverses fragile X syndrome-like phenotypes. Cell Rep 9: 1742-1755
- 81. Zybura-Broda K, Wolder-Gontarek M, Ambrozek-Latecka M, Choros A, Bogusz A, Wilemska-Dziaduszycka J, Rylski M. (2018) HuR (Elavl1) and HuB (Elavl2) Stabilize matrix metalloproteinase-9 mRNA during seizure-induced Mmp-9 expression in neurons. Front Neurosci 12: 224
- Dziembowska M, Włodarczyk J (2012) MMP9: a novel function in synaptic plasticity. Int J Biochem Cell Biol 44: 709-713
- 83. Stawarski M, Stefaniuk M, Wlodarczyk J (2014) Matrix metalloproteinase-9 involvement in the structural plasticity of dendritic spines. Front Neuroanat 8: 68
- 84. Bilousova TV, Dansie L, Ngo M, Aye J, Charles JR, Ethell DW, Ethell IM (2009) Minocycline promotes dendritic spine maturation and improves behavioural performance in the fragile X mouse model. J Med Genet 46: 94-102
- 85. Michaluk P, Wawrzyniak M, Alot P, Szczot M, Wyrembek P, Mercik K, Medvedev N, Wilczek E, De Roo M, Zuschratter W, Muller D, Wilczynski GM, Mozrzymas JW, Stewart MG, Kaczmarek L, Wlodarczyk J (2011) Influence of matrix metalloproteinase MMP-9 on dendritic spine morphology. J Cell Sci 124: 3369-3380
- 86. Wang XB, Bozdagi O, Nikitczuk JS, Zhai ZW, Zhou Q, Huntley GW (2008) Extracellular proteolysis by matrix metalloproteinase-9 drives dendritic spine enlargement and long-term potentiation coordinately. Proc Natl Acad Sci USA 105: 19520-19525
- Magnowska M, Gorkiewicz T, Suska A, Wawrzyniak M, Rutkowska-Wlodarczyk I, Kaczmarek L, Wlodarczyk J (2016) Transient ECM protease activity promotes synaptic plasticity. Sci Rep 6: 27757
- 88. Sidhu H, Dansie LE, Hickmott PW, Ethell DW, Ethell IM (2014) Genetic removal of matrix metalloproteinase 9 rescues the symptoms of fragile X syndrome in a mouse model. J Neurosci 34: 9867-9879
- Aujla PK, Huntley GW (2014) Early postnatal expression and localization of matrix metalloproteinases-2 and -9 during establishment of rat hippocampal synaptic circuitry. J Comp Neurol 522: 1249-1263
- 90. Murase S, Lantz CL, Kim E, Gupta N, Higgins R, Stopfer M, Hoffman DA, Quinlan EM (2016) Matrix metalloproteinase-9 regulates neuronal circuit development and excitability. Mol Neurobiol 53: 3477-3493
- 91. Fragkouli A, Papatheodoropoulos C, Georgopoulos S, Stamatakis A, Stylianopoulou F, Tsilibary EC, Tzinia AK (2012) Enhanced neuronal plasticity and elevated endogenous sAPPalpha levels in mice overexpressing MMP9. J Neurochem 121: 239-251
- 92. Tsien RY (2013) Very long-term memories may be stored in the pattern of holes in the perineuronal net. Proc Natl Acad Sci USA 110: 12456-12461
- 93. Frischknecht R, Heine M, Perrais D, Seidenbecher CI, Choquet D, Gundelfinger ED (2009) Brain extracellular matrix affects AMPA receptor lateral mobility and short-term synaptic plasticity. Nat Neurosci 12: 897-904
- 94. Dityatev A, Rusakov DA (2011) Molecular signals of plasticity at the tetrapartite synapse. Curr Opin Neurobiol 21: 353-359
- 95. Soleman S, Filippov MA, Dityatev A, Fawcett JW (2013) Targeting the neural extracellular matrix in neurological disorders. Neuroscience 253: 194-213
- 96. Tamura H, Ishikawa Y, Shiosaka S (2013) Does extracellular proteolysis control mammalian cognition? Rev Neurosci. 24: 365-374
- 97. Foscarin S, Ponchione D, Pajaj E, Leto K, Gawlak M, Wilczynski GM, Rossi F, Carulli D (2011) Experience-dependent plasticity and modulation of growth regulatory molecules at central synapses. PLoS One 6: e16666
- 98. Stamenkovic V, Stamenkovic S, Jaworski T, Gawlak M, Jovanovic M, Jakovcevski I, Wilczynski GM, Kaczmarek L, Schachner M, Radenovic L, Andjus PR (2017) The extracellular matrix glycoprotein Tenascin-C and matrix metalloproteinases modify cerebellar structural plasticity

by exposure to an enriched environment brain structure and function. Brain Struct Funct 222: 393-415

- 99. Bijata M, Labus J, Guseva D, Stawarski M, Butzlaff M, Dzwonek J, Schneeberg J, Böhm K, Michaluk P, Rusakov DA, Dityatev A, Wilczyński G, Wlodarczyk J, Ponimaskin E (2017) Synaptic remodeling depends on signaling between serotonin receptors and the extracellular matrix. Cell Rep 19: 1767-1782
- 100. Michaluk P, Mikasova L, Groc L, Frischknecht R, Choquet D, Kaczmarek L (2009) Matrix metalloproteinase-9 controls NMDA receptor surface diffusion through integrin beta1 signaling. J Neurosci 29: 6007-6012
- Bajor M, Kaczmarek L (2013) Proteolytic Remodeling of the Synaptic Cell Adhesion Molecules (CAMs) by metzincins in synaptic plasticity. Neurochem Res 38: 1113-1121
- 102. Conant K, Allen M, Lim ST (2015) Activity Dependent CAM cleavage and Neurotransmission. Front Cell Neurosci 9: 305
- 103. Shinoe T, Goda Y (2015) Tuning synapses by proteolytic remodeling of the adhesive surface. Curr Opin Neurobiol 35: 148-155
- 104. Tian L, Stefanidakis M, Ning L, Van Lint P, Nyman-Huttunen H, Libert C, Itohara S, Mishina M, Rauvala H, Gahmberg CG (2007) Activation of NMDA receptors promotes dendritic spine development through MMP-mediated ICAM-5 cleavage. J Cell Biol 178: 687-700
- 106. Kelly EA, Tremblay ME, Gahmberg CG, Tian L, Majewska AK (2014) Subcellular localization of intercellular adhesion molecule-5 (Telencephalin) in the visual cortex is not developmentally regulated in the absence of Matrix Metalloproteinase-9. J Comp Neurol 522: 676-688
- 107. Peixoto RT, Kunz PA, Kwon H, Mabb AM, Sabatini BL, Philpot BD, Ehlers MD (2012) Transsynaptic signaling by activity-dependent cleavage of Neuroligin-1. Neuron 76: 667-667
- 107. van der Kooij M, Fantin M, Rejmak E, Grosse J, Zanoletti O, Fournier C, Ganguly K, Kalita K, Kaczmarek L, Sandi C (2014) Role for MMP-9 in stress-induced downregulation of nectin-3 in hippocampal CA1 and associated behavioural alterations. Nat Commun 5: 4995
- 108. Stawarski M, Rutkowska-Wlodarczyk I, Zeug A, Bijata M, Madej H, Kaczmarek L, Wlodarczyk J (2014) Genetically encoded FRET-based biosensor for imaging MMP-9 activity. Biomaterials 35: 1402-1410
- 109. Bajor M, Michaluk P, Gulyassy P, Kekesi AK, Juhasz G, Kaczmarek L (2012) Synaptic cell adhesion molecule-2 and collapsin response mediator protein-2 are novel members of the matrix metalloprotein-ase-9 degradome. J Neurochem 122: 775-788
- 110. Bruno MA, Cuello AC (2006) Activity-dependent release of precursor nerve growth factor, conversion to mature nerve growth factor, and its degradation by a protease cascade. Proc Natl Acad Sci USA 103: 6735-6740
- 111. Mizoguchi H, Nakade J, Tachibana M, Ibi D, Someya E, Koike H, Kamei H, Nabeshima T, Itohara S, Takuma K, Sawada M, Sato J, Yamada K (2011) Matrix metalloproteinase-9 contributes to kindled seizure development in pentylenetetrazole-treated mice by converting pro-BDNF to mature BDNF in the hippocampus. J Neurosci 31: 12963-12971
- 112. Reinhard SM, Razak K, Ethell I (2015) A delicate balance: role of MMP-9 in brain development and pathophysiology of neurodevelopmental disorders. Front Cell Neurosci 9: 280
- 113. Lukasiuk K, Wilczynski GM, Kaczmarek L (2011) Extracellular proteases in epilepsy. Epilepsy Res 96: 191-206
- 114. Kim GW, Kim HJ, Cho KJ, Kim HW, Cho YJ, Lee BI (2009) The role of MMP-9 in integrin-mediated hippocampal cell death after pilocarpine-induced status epilepticus. Neurobiol Dis 36: 169-180
- 115. Takacs E, Nyilas R, Szepesi Z, Baracskay P, Karlsen B, Røsvold T, Bjørkum AA, Czurkó A, Kovács Z, Kékesi AK, Juhász G (2010) Matrix metalloproteinase-9 activity increased by two different types of epileptic seizures that do not induce neuronal death: a possible role in homeostatic synaptic plasticity. Neurochem Int 56: 799-809
- 116. Yeghiazaryan M, Rutkowska-Wlodarczyk I, Konopka A, Wilczynski GM, Melikyan A, Korkotian E, Kaczmarek L, Figiel I (2014) DP-b99 modulates matrix metalloproteinase activity and neuronal plasticity. PLoS ONE 9: e99789

- 117. Pijet B, Stefaniuk M, Kostrzewska-Ksiezyk A, Tsilibary P-E, Tzinia A, Kaczmarek L Elevation of MMP-9 levels promotes epileptogenesis after traumatic brain injury. Mol. Neurobiol., in press.
- 118. Li S, Yu SX, Zhang CQ, Shu HF, Liu SY, An N, Yang MH, Yin Q, Yang H (2012) Increased expression of matrix metalloproteinase 9 in cortical lesions from patients with focal cortical dysplasia type IIb and tuberous sclerosis complex. Brain Res 1453: 46-55
- 119. Knapska E, Lioudyno V, Kiryk A, Gorkiewicz T, Mikosz M, Michaluk P, Gawlak M, Chaturvedi M, Mochol G, Balcerzyk M, Wojcik DK, Wilczynski GM, Kaczmarek L (2013) Reward learning requires activity of matrix metalloproteinase-9 in the central amygdala. J Neurosci 33: 14591-14600
- 120. Quirico-Santos T, Nascimento Mello A, Casimiro Gomes A, de Carvalho LP, de Souza JM, Alves-Leon S (2013) Increased metalloprotease activity in the epileptogenic lesion -Lobectomy reduces metalloprotease activity and urokinase-type uPAR circulating levels. Brain Res 1538: 172-181
- 121. Zybura-Broda K, Amborska R, Ambrozek-Latecka M, Wilemska J, Bogusz A, Bucko J, Konopka A, Grajkowska W, Roszkowski M, Marchel A, Rysz A, Koperski L, Wilczynski GM, Kaczmarek L, Rylski M (2016) Epigenetics of Epileptogenesis-Evoked Upregulation of Matrix Metalloproteinase-9 in Hippocampus. PLoS One 11: e0159745
- 122. Smith AC, Kupchik YM, Scofield MD, Gipson CD, Wiggins A, Thomas CA, Kalivas PW (2014) Synaptic plasticity mediating cocaine relapse requires matrix metalloproteinases. Nat Neurosci 17: 1655-1657
- Mulholland PJ, Chandler LJ, Kalivas PW (2016) Signals from the fourth dimension regulate drug relapse. Trends Neurosci 39: 472-485
- 124. Mash DC, Ffrench-Mullen J, Adi N, Qin YJ, Buck A, Pablo J (2007) Gene expression in human hippocampus from cocaine abusers identifies genes which regulate extracellular matrix remodeling. Plos One 2: e1187
- 125. Brown TE, Forquer MR, Cocking DL, Jansen HT, Harding JW, Sorg BA (2007) Role of matrix metalloproteinases in the acquisition and reconsolidation of cocaine-induced conditioned place preference. Learn Mem 14: 214-223
- 126. Brown TE, Forquer MR, Harding JW, Wright JW, Sorg BA (2008) Increase in matrix metalloproteinase-9 levels in the rat medial prefrontal cortex after cocaine reinstatement of conditioned place preference. Synapse 62: 886-889
- 127. Conant K, Lonskaya I, Szklarczyk A, Krall C, Steiner J, Maguire-Zeiss K, Lim ST (2011) Methamphetamine-associated cleavage of the synaptic adhesion molecule intercellular adhesion molecule-5. J Neurochem 118: 521-532
- 128. Samochowiec A, Grzywacz A, Kaczmarek L, Bienkowski P, Samochowiec J, Mierzejewski P, Preuss UW, Grochans E, Ciechanowicz A (2010) Functional polymorphism of matrix metalloproteinase-9 (MMP-9) gene in alcohol dependence: family and case control study. Brain Res 1327: 103-106
- 129. Mizoguchi H, Yamada K, Mouri A, Niwa M, Mizuno T, Noda Y, Nitta A, Itohara S, Banno Y, Nabeshima T (2007) Role of matrix metalloproteinase and tissue inhibitor of MMP in methamphetamine-induced behavioral sensitization and reward: implications for dopamine receptor down-regulation and dopamine release. J Neurochem 102: 1548-1560
- 130. Mizoguchi H, Yamada K, Mouri A, Niwa M, Mizuno T, Noda Y, Nitta A, Itohara S, Banno Y, Nabeshima T (2007) Reduction of methamphetamine-induced sensitization and reward in matrix metalloproteinase-2 and-9-deficient mice. J Neurochem 100: 1579-1588
- 131. Spencer S, Neuhofer D, Chioma VC, Garcia-Keller C, Schwartz DJ, Allen N, Scofield MD, Ortiz-Ithier T, Kalivas PW. A model of Δ9-tetrahydrocannabinol self-administration and reinstatement that alters synaptic plasticity in nucleus accumbens. Biol Psychiatry, in press
- 132. Stefaniuk M, Beroun A, Lebitko T, Markina O, Leski S, Meyza K, Grzywacz A, Samochowiec J, Samochowiec A, Radwanska K, Kaczmarek L (2017) Matrix metalloproteinase-9 and synaptic plasticity in the central amygdala in control of alcohol seeking behavior. Biol Psychiat, 81: 905–906

- 133. Rotschafer SE, Trujillo MS, Dansie LE, Ethell IM, Razak KA (2012) Minocycline treatment reverses ultrasonic vocalization production deficit in a mouse model of Fragile X Syndrome. Brain Res 1439: 7-14
- 134. Wen TH, Afroz S, Reinhard SM, Palacios AR, Tapia K, Binder DK, Razak KA, Ethell IM (2017) Genetic reduction of matrix metalloproteinase-9 promotes formation of perineuronal nets around parvalbumin-expressing interneurons and normalizes auditory cortex responses in developing Fmr1 knock-out mice. Cereb Cortex 13: 1-14
- 135. Paribello C, Tao L, Folino A, Berry-Kravis E, Tranfaglia M, Ethell IM, Ethell DW (2010) Open-label add-on treatment trial of minocycline in fragile X syndrome. BMC Neurol 10: 91
- 136. Leigh MJS, Nguyen DV, Mu Y, Winarni TI, Schneider A, Chechi T, Polussa J, Doucet P, Tassone F, Rivera SM, Hessl D, Hagerman RJ (2013) A randomized double-blind, placebo-controlled trial of minocycline in children and adolescents with fragile x syndrome. J Dev Behav Pediatr 34: 147-155
- 137. Siller SS, Broadie K (2012) Matrix metalloproteinases and minocycline: therapeutic avenues for fragile X syndrome. Neural Plast 2012: 124548

- 138. Dziembowska M, Pretto DI, Janusz A, Kaczmarek L, Leigh MJ, Gabriel N, Durbin-Johnson B, Hagerman RJ, Tassone F (2013) High MMP-9 activity levels in fragile X syndrome are lowered by minocycline. Am J Med Genet A 161A: 1897-1903
- Hagerman RJ, Polussa J (2015) Treatment of the psychiatric problems associated with fragile X syndrome. Curr Opin Psychiatry 28: 107-112
- Lepeta K, Kaczmarek L (2015) Matrix Metalloproteinase-9 as a novel player in synaptic plasticity and schizophrenia. Schizophr Bull 41: 1003-1009
- 142. Lepeta K, Purzycka K, Pachulska-Wieczorek K, Mitjans M, Begemann M, Vafadari B, Bijata K, Adamiak R, Ehrenreich H, Dziembowska M, Kaczmarek L (2017) A normal genetic variation modulates synaptic MMP-9 protein levels and the severity of schizophrenia symptoms. EMBO Mol Med 9: 1100-1116

# Od c-Fos do MMP-9 w kontroli plastyczności synaptycznej zdrowego i chorego umysłu, spojrzenie osobiste

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Słowa kluczowe: uczenie się i pamięć, cząsteczki adhezji komórkowej, padaczka, autyzm, schizofrenia, uzależnienia

#### STRESZCZENIE

c-Fos jest składnikiem czynnika transkrypcyjnego AP-1. Trzy linie dowodów wspierają kluczową rolę c-Fos w uczeniu się i pamięci: (i) uczenie się pobudza jego ekspresję; (ii) blokowanie c-Fos upośledza, podczas gdy optogenetyczna aktywacja neuronów wyrażających c-Fos wspomaga uczenie się i pamięć; (iii) docelowe geny dla c-Fos/AP-1 w pobudzonych neuronach, kodują tkankowy inhibitor metaloproteaz-1 (TIMP-1) oraz metaloproteazę macierzową 9 (MMP-9), które to białka odgrywają kluczową rolę w plastyczności synaptycznej, leżącej u podstaw uczenia się i pamięci. TIMP-1 i MMP-9 tworzą działający zewnątrzkomórkowo układ enzymatyczny, aktywny miejscowo wokół synaps pobudzających i modulujący ich morfologię, skład biochemiczny oraz efektywność. Badania na zwierzętach sugerują zaangażowanie MMP-9 w różnych stanach neuropsychicznych, np. rozwoju padaczki, zaburzeniach ze spektrum autyzmu, rozwoju uzależnienia i depresji. U ludzi MMP-9 przyczynia się do padaczki, uzależnienia od alkoholu i kokainy, zespołu łamliwego chromosomu X, schizofrenii i zaburzeń afektywnych dwubiegunowych. Łącznie wszystkie te sytuacje można uznać za przykładowe dla zdrowego lub chorego umysłu.